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TITLE: AKT Rescue in Cardiomyocytes but not Breast Cancer Cells after Doxorubicin and Anti-erbB2 Treatment

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## Abstract

**Study Design:** The proposed study will first evaluate the role of Akt, in protection against doxorubicin and anti-erbB2- cardiomyocyte toxicity, using adenoviral expression of active Akt, pharmacological inhibitors of this pathway, and two peptides that activate Akt, cardiotrophin-1 and urocortin. Since these peptides have not been reported expressed in breast tissues or cancer, to confirm this, we will evaluate the expression of both peptides and their receptors in six commonly studied breast cancer cell lines and 160 breast cancer tissue arrays by immuno-histochemistry and western blotting methods. Even if expression is observed in breast tissue, peptide treatment may improve cancer therapy as seen in other models. In aim 3, the cardiotrophin-1 and urocortin cardiac protection strategy, will be tested against cardiac toxicity induced by doxorubicin, anti-erbB2, chemical inhibitors of erbB1 or erbB2, or combination treatments. This will be a direct comparison of rat and human cardiomyocytes with 6 breast cancer cell lines using MTT assay. Next both peptides, will be administered in pilot studies to Sprague Dawley rats to establish a dose that protects against doxorubicin induced cardiac toxicity. Finally, using a female nude rat breast cancer xenograph model, these peptides will be evaluated for specific cardiac protection, during treatment with doxorubicin, anti-erbB2, combination of doxorubicin and anti-erbB2 and controls. Echocardiography, to evaluate ejection fraction, white blood cell counts, to evaluate bone marrow toxicity, histopathology, xenograph tumor size and weights will be used to assess peptide cardiac specific protection and anti-neoplastic therapy.

**Relevance:** Doxorubicin is currently a first choice drug for breast cancer treatment, limited in use by its cardiac toxicity. Combination drug treatment is the standard of care. This proposal addresses a timely clinical problem observed with doxorubicin and anti-erbB2 combined therapy, as well as the potential problem of combined treatment with doxorubicin with other erbB inhibitors. This strength of this proposal is the direct comparison of relevant human cardiomyocytes and human breast cancer cells to better understand the mechanism of toxicity and to evaluate in human cells, a novel peptide protection strategy aimed to reduce cardiotoxicity without affecting breast cancer anti-neoplastic therapy.

## Introduction

### Significance of the project:

Anti-erbB2 (Herceptin) and doxorubicin are effective treatments for breast cancers that over-express the Her2/neu oncogene. In pivotal trials that lead to its approval, anti-erbB2 administered in combination with other agents, particularly doxorubicin, 29% of patients developed cardiac dysfunction, and in some cases fatal, cardiomyopathy [1-3, 10]. The mechanism of this synergistic toxicity induced by anti-erb2 is not understood. To address this problem, we have developed novel *in vitro* and *in vivo* rat animal models that exhibit cardiac toxicity synergism with doxorubicin and anti-rat erbB2 compared to either treatment alone. We utilize an anti-rat-erbB2 monoclonal antibody (clone 7.16.4) that shows the same biological effects in rat cells expressing erbB2, as Herceptin does in human breast cancer cells, including similar epitope recognition, inhibition of cell growth, reversion of phenotype and reduction in cancer cell growth *in vivo* [4]. In this model, Akt, a well-known anti-apoptotic pathway protein in the heart, linked to the erbB2, is inactivated. This finding of Akt inactivation is consistent with Herceptin's mechanism of action in breast cancer cells [5, 6]. Cardiotrophin-1 and urocortin both stimulate the Akt pathway through different receptors expressed in the heart [7-9], and not breast cells; thus, treatment with these peptides, may circumvent the erbB-linked Akt pathway and provide protection during doxorubicin or doxorubicin/anti-erbB2 treatment.

**The hypothesis of this proposal is that activation of Akt in the heart through heart specific receptors, during doxorubicin and anti-erbB2 therapy, will protect from cardiac toxicity and not diminish doxorubicin and anti-erbB2 tumor cell killing in breast cancer cells.**

**Specific Aim 1** Determine the role of Akt activation, by non-erbB2 pathways, in protection of cardiomyocytes against toxicity induced by anti-erbB2, doxorubicin, chemical inhibitors of erbB1 or erbB2 or combination treatments.

**Specific Aim 2** Screen a panel of commonly used human breast cancer cell lines, breast cancer tissue arrays and normal epithelium arrays for expression to urocortin or cardiotrophin-1 peptides or their receptors.

**Specific Aim 3** Determine whether activation of Akt, by the non-erbB2 pathways, (urocortin or cardiotrophin-1), will preferentially protect cardiomyocytes and not breast cancer cells, against toxicity induced by anti-erbB2, doxorubicin, inhibitors of erbB1 or erbB2 or combined treatments.

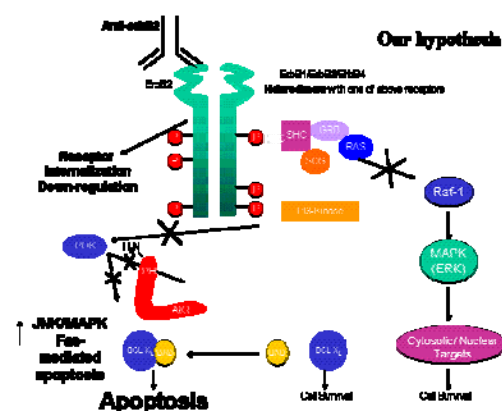
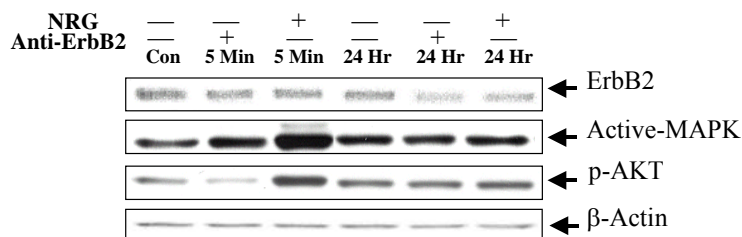
**Specific Aim 4** Determine whether *in vivo* pretreatment with Akt-inducing urocortin or cardiotrophin-1 provides protection for doxorubicin and /or anti-erbB2 treatment induced cardiac toxicity in rats without affecting anti-neoplastic effects of therapy.

### Body: Research Accomplishments

To test our hypothesis, our specific aims are:

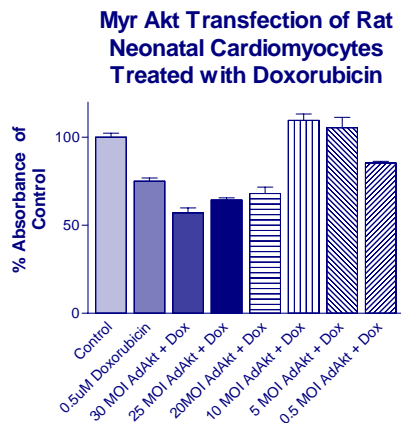
**Specific Aim 1** Determine the role of Akt activation, by non-erbB2 pathways, in protection of cardiomyocytes against toxicity induced by anti-erbB2, doxorubicin, chemical inhibitors of erbB1 or erbB2 or combination treatments.

### Anti-erbB2 reduces activation of Akt and ERK1/2MAPK compared to NRG

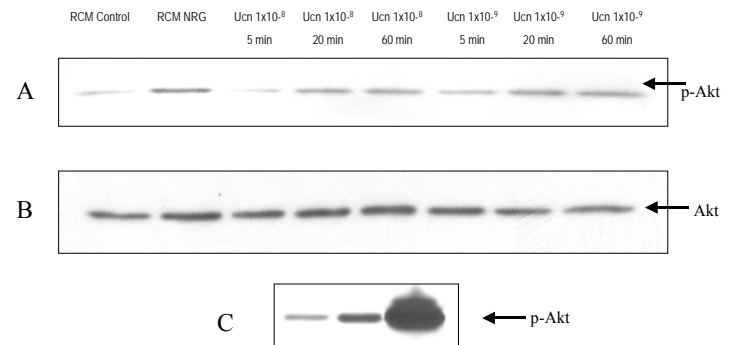


An Akt expressing adenovirus induced protection against doxorubicin. MOI of 5-10 showed the best protection (MTT assay below left). Below right (A) Western blot from urocortin-treated cardiomyocytes lysates. The

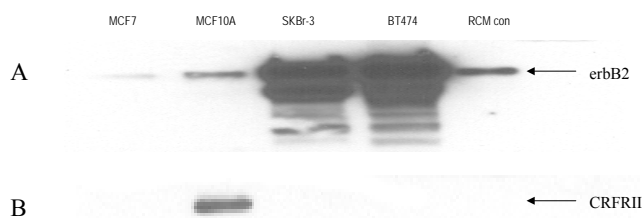
amount of phosphorylated Akt increases as urocortin concentration and exposure time increase. (B) The amount of Akt stays the same as urocortin concentration and exposure time increase. (C) When cardiomyocyte lysates were treated with an adenovirus that expresses active Akt, the amount of Akt in the cells increased lane 3, lane 2 NRG treatment, lane 1 control.



Cardiomyocytes treated with an adenovirus that expresses active Akt are protected from cardiac toxicity. (n=12)



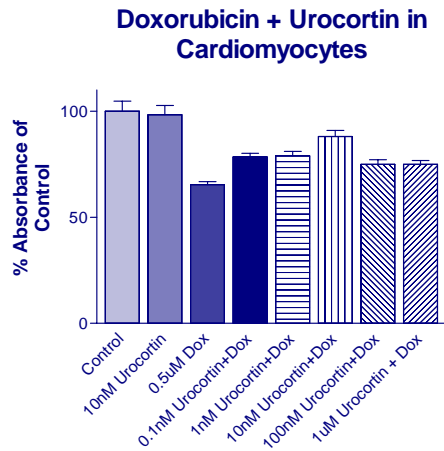
**Specific Aim 2** Screen a panel of commonly used human breast cancer cell lines, breast cancer tissue arrays and normal epithelium arrays for expression to urocortin or cardiotrophin-1 peptides or their receptors.



Cell Lines	Urocortin	CRFR II	ErbB2
BT474	No expression	No expression	High expression
MCF7	No expression	No expression	Low expression
MCF10A	No expression	Moderate expression	Low expression
SKBr-3	No expression	No expression	High expression

The previous blot compares protein expression between RCM (rat neonatal cardiomyocytes, control) and various breast cancer cell lines. The antibody to CRFR II only cross reacts with the human protein and not the rat protein. At present, there is not a suitable antibody for the rat. The expression of urocortin and its receptor, CRF2 are summarized in the above table. It is known that urocortin does protect cardiomyocytes from ischemia/reperfusion conditions. Since urocortin treatment offered protection to cardiomyocytes, we screened a panel of commonly used human breast cancer cell lines for the expression of urocortin and its receptor, CRFR II. We next determined whether activation of Akt by the non-erbB2 pathway, urocortin, would preferentially protect cardiomyocytes and not breast cancer cells against toxicity induced by doxorubicin. Results are presented below.

**Specific Aim 3** Determine whether activation of Akt, by the non-erbB2 pathways, (urocortin or cardiotropin-1), will preferentially protect cardiomyocytes and not breast cancer cells, against toxicity induced by anti-erbB2, doxorubicin, inhibitors of erbB1 or erbB2 or combined treatments.



Above Left figure: The number of metabolically active neonatal rat cardiomyocytes was determined by the MTT assay. Neonatal rat cardiomyocytes were treated doxorubicin and urocortin. Urocortin protects neonatal rat cardiomyocytes from cardiac toxicity. (n=36)

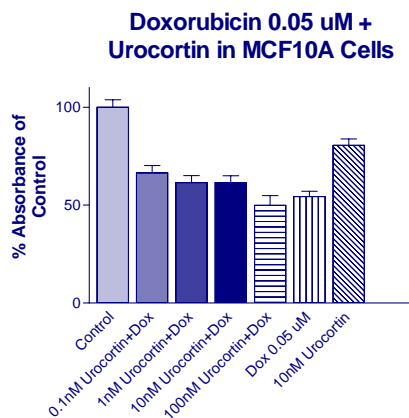


Figure (left) The number of metabolically active MCF10A cells was determined by the MTT assay. MCF10A cells were treated Doxorubicin (0.05uM) and Urocortin (variable concentrations). Urocortin does not protect MCF10A cells from doxorubicin toxicity. (n=12)  
This suggests that stimulation of other Akt pathways, may be protective with dual doxorubicin and anti-erbB2 treatment in the heart but not in breast cells that have a receptor. Future studies will test if urocortin can offer protection in this combined treatment in cardiomyocytes.



Figure 10: Western blots from urocortin treated MCF10A (20 min). The amount of phosphorylated Akt in the cells does not change as urocortin concentration increases. RRM=rat neonatal cardiomyocytes, NRG= neuregulin 1 $\beta$ , positive control for Akt activation

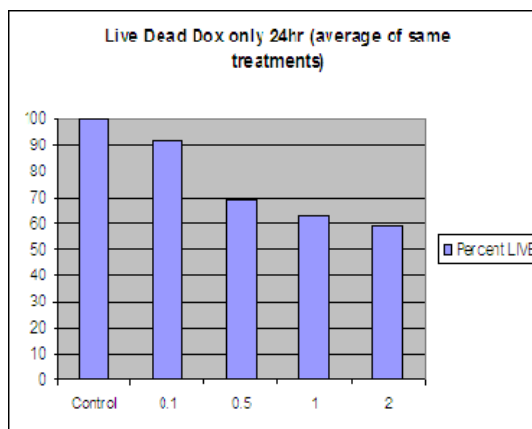
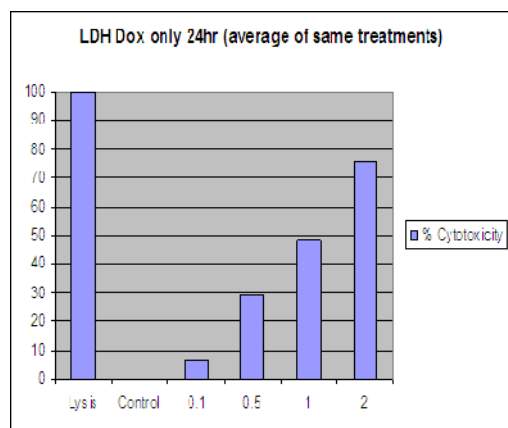
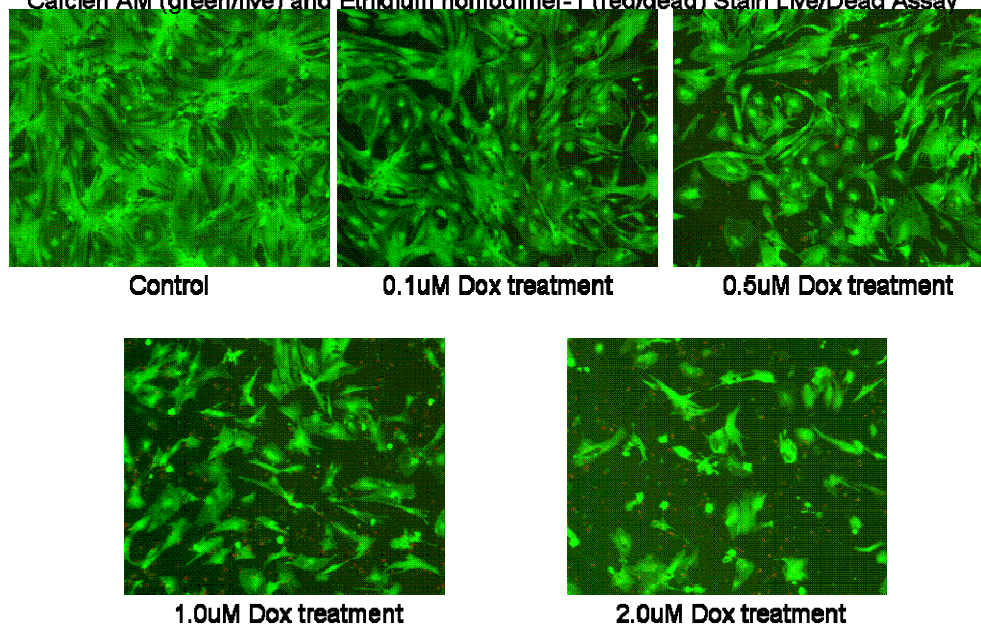


Figure 11: Western blots from MCF10A (20 min) treated with Urocortin (10 nM) for varying times. The amount of phosphorylated Akt in the cells does not change as exposure time increases. RRM=rat neonatal cardiomyocytes, NRG= neuregulin 1 $\beta$ , positive control for Akt activation

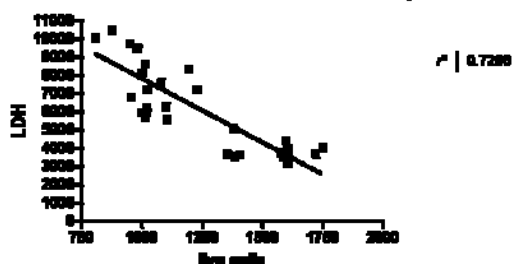
The following is validation of two cytotoxicity assays with doxorubicin which will apply to Aim 1 and 3: These two assays will be used in doxorubicin and erbB2 inhibition, as well as PI3K inhibition experiments. These assays will also be necessary to use in doxorubicin toxicity prevention experiments. Our prior experiments used the MTT assay which measures mitochondrial function and not always correlated with cell death (multiple reviewer's comments). We will next analyze how MTT, LDH and the Live/Dead cell assays correlate by doing linear regression statistics, as was done with the validation of LDH and the Live/Dead cell assay compared in the below figure.

## Dose/Response

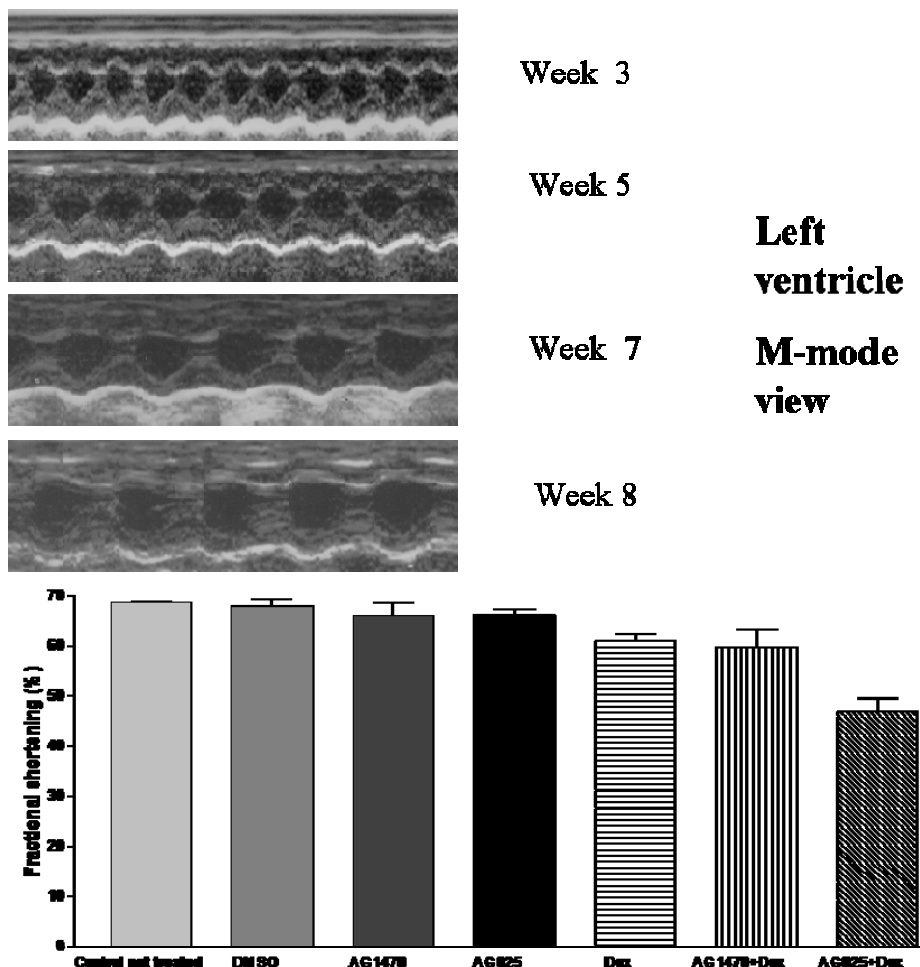
Calcein AM (green/live) and Ethidium homodimer-1 (red/dead) Stain Live/Dead Assay



## Correlation - LDH and Live/Dead cell assays



This year we also developed a mouse model of chronic doxorubicin toxicity. Below is the M-mode of the left ventricle over time. This figure shows the progressive dilation and loss of cardiac contractility with administration of (4) tail vein intravenous injections of 9 mg/kg doxorubicin given every 2 weeks.



Using this same model, but using only 3 tail vein intravenous injections of 9 mg/kg doxorubicin given every 2 weeks, we applied the concept in Specific Aim 4 by inhibiting erbB1 and erbB2. Giving a combined treatment of Doxorubicin and AG825, an erbB2 inhibitor, synergistic toxicity was observed resulting in a marked reduction in the cardiac fractional shortening evaluated by in vivo transthoracic echocardiography. This model will be evaluated for protection with urocortin, an pAKT inducer along side the rat model.

**Monoclonal antibody production:** We require a large amount of antibody to perform the anti-erbB2 in vitro and in vivo experiments. We had a three month period when we could not make antibody because of a technical problem that was finally solved. This had an impact on rat studies, although the mouse model listed above may be added if we can not make enough antibody for these complete studies. I will see what can be accomplished this year before changing the protocol. The hybridoma we use to produce 7.16.4 has always been grown in an Integra flask (Integra Biosciences). We have two flasks going at a time to make about 5mg/week. In order to meet the large quantity of antibody we need for these animal studies, we also set up a Hollow fiber system HFS (Bellco) to increase antibody production. Our antibody is routinely checked for activity by flow cytometry and western blotting (erbB2 phosphorylation). Each rat in a study requires 4mg/study. We were unsuccessful in adapting our hybridoma to the HFS, even with close consultation with the company's president. Some hybridomas are not adaptable to the HFS.

### Key Research Accomplishments

- Akt expressing Adenovirus protects against doxorubicin toxicity in cardiomyocytes
- Urocortin induces AKT activation (pAKT) in cardiomyocytes
- Comparison of erbB2 and CRF2 expression in breast cancer cell lines and cardiomyocytes
- Only MCF10A cells have CRF2 receptor for urocortin
- Urocortin provides protection against doxorubicin toxicity in cardiomyocytes
- Urocortin does not provides protection against doxorubicin toxicity in MCF10A cells
- Validation of two cytotoxicity assays for this project
- Inhibition of erbB2 in a mouse model of doxorubicin toxicity induces synergistic cardiac toxicity

### Reportable Outcomes



## Manuscripts

**Gabrielson KL**, Becker R, Shi W, Servinsky M, Bedja D, Barber S, Akao M, Peterson N. Synergistic cardiac toxicity with doxorubicin and anti-erbB2 treatment in rats: Model for Herceptin-induced cardiac toxicity. In revision

## Presentations

JHU- Department of Oncology Breast cancer research SPOR program July 2004, "Breast cancer therapies that induce cardiac toxicity- Can cardiac Akt activation prevent?"

JHU-ICMIC P50 program Department of Radiology September 2004, "Role of Akt pathway in immuno and chemotherapy for breast cancer"

University of Colorado, Denver, School of Medicine, Department of Oncology "Synergistic toxicity induced by doxorubicin and anti-erbB2 in rat model", January 2005

## Conclusions

**In the first year of this project, we have accomplished several of the proposed experiments in the Statement of Work in Aims 1-3. Activated AKT is protective during doxorubicin toxicity in cardiomyocytes. We identified a peptide (urocortin) that also provides protection during doxorubicin toxicity in cardiomyocytes. Most importantly, we screened several breast cancer cell lines and did not find expression of the receptor for this peptide. Thus, this peptide is a candidate to test in vivo for protection against doxorubicin and the combined anti-erbB2/doxorubicin treatment.**

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## Summary Part I

### Validation of the Use of Long-Term Indwelling Jugular Catheters in a Rat Model of Cardiac Toxicity

Doxorubicin administered to rats induces a dose-dependent cardiomyopathy creating a useful in-vivo model for investigation of cardiotoxicity. Both doxorubicin administration and presence of indwelling venous catheters have been associated with thrombus formation. The study aim was to determine feasibility of drug delivery and degree of thrombogenesis related to long-term indwelling catheter use in a cardiotoxicity model. Rats receiving doxorubicin or saline via jugular catheters coated with end-point immobilized heparin were compared to rats receiving similar treatments via direct jugular intravenous injection (venotomy). Onset of cardiotoxicity, defined by reduction in fractional shortening to or below 45%, was determined by echocardiography. Thrombogenesis was assessed by observation of atrial thrombi and pulmonary emboli as determined by post-mortem and histologic examination. A significantly greater proportion of the doxorubicin treated/catheterized group (87.5%) achieved cardiotoxicity relative to the doxorubicin treated/venotomy group (28.6%). This was reflected by an earlier and more precipitous decline in fractional shortening demonstrated by the doxorubicin treated/catheterized rats. Despite this, rats from catheterized groups demonstrated improved weight maintenance relative to venotomy groups. Although significant differences in presence of pulmonary emboli between groups were not observed, 50% of the doxorubicin treated/catheterized group developed vegetative endocarditis. Although development of such lesions represents alteration of the model-induced cardiac disease, we submit that the more reliable and early induction of desired endpoint, in addition to improved weight maintenance, represent model refinements. Development of valvular vegetations is similar to complications observed in cancer patients such that this model may serve as a means to study formation and prevention of such lesions.

**This study determined that we could use the catheterized rat for this model. Using this system will allow a central line in the rat that can be used to deliver inhibitors of erbB2 pathway (antibody and small molecule inhibitors) as well as withdraw blood that can be tested for cardiac biomarkers of damage. The below tables and figures are part of a manuscript in press.**

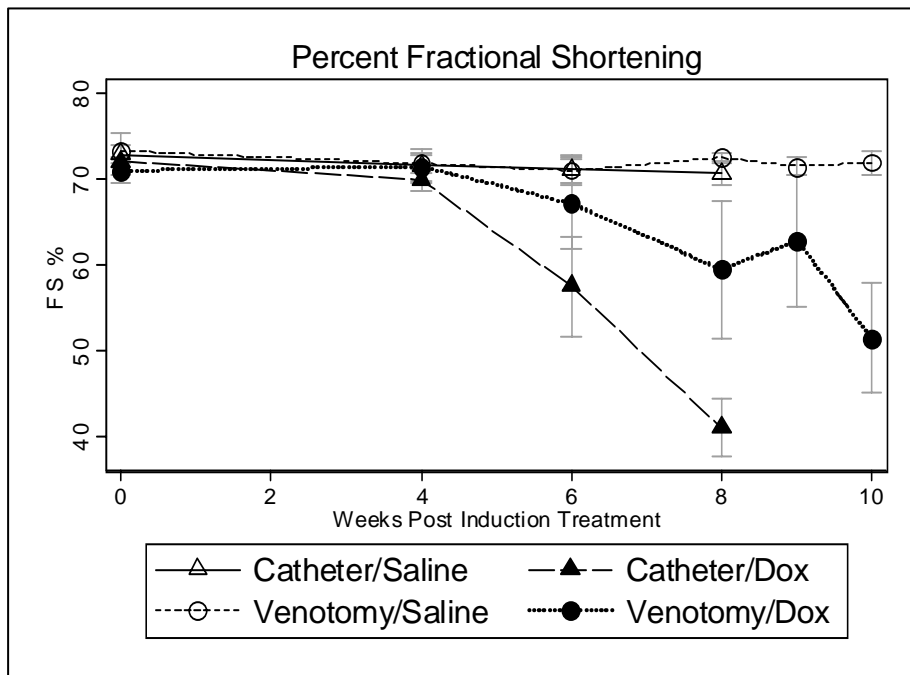
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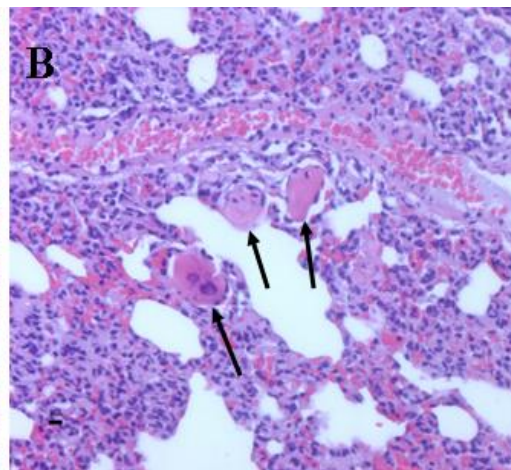
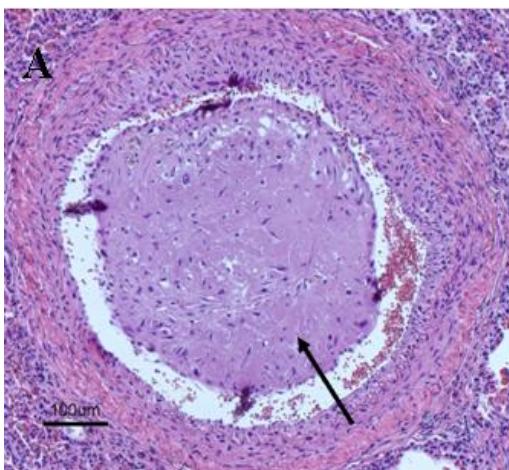
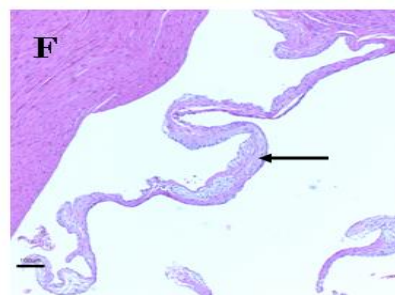
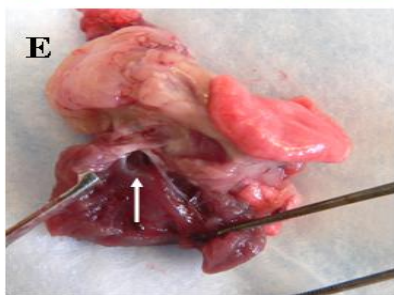
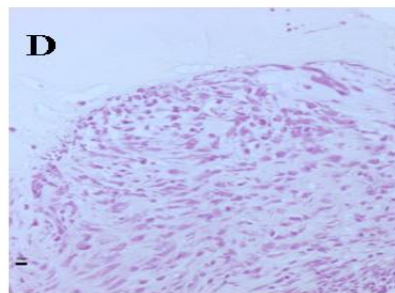
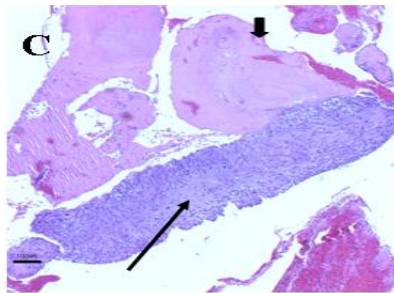
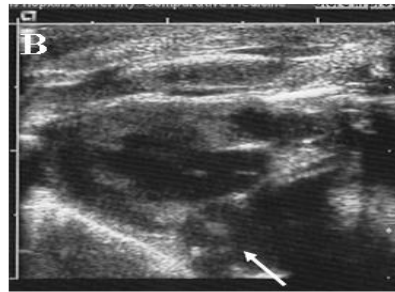
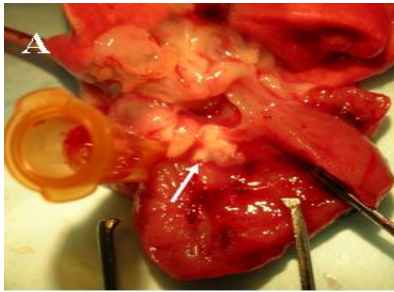
<i>Rat / Treatment group</i>	<i>Vegetation/ location</i>	<i>Pulmonary emboli</i>	<i>Atrial thrombi</i>	<i>Catheter tract contamination</i>
<b><i>Group A: Doxorubicin treated/Catheterized</i></b>				
Rat A	+ LAV	—	+	+ <sup>†</sup>
Rat B	+ LAV	+	+	—
Rat C	+ RAV	—	—	—
Rat D	+ RAV	—	—	+
Rat E	—	+	—	+
Rat F	—	+*	—	—
Rats G-H	—	—	—	—
<b><i>Group B: Saline treated/Catheterized</i></b>				
Rat A	—	+	—	—
Rats B-H	—	—	—	—
<b><i>Group C: Doxorubicin treated/Venotomy</i></b>				
Rat A	—	+	+	NA
Rats B-G	—	—	—	NA
<b><i>GroupD: Saline treated/Venotomy</i></b>				
Rats A-G	—	—	—	NA
<b>Fisher Exact P value</b>	<b>&lt;0.01</b>	NS	NS	<b>&lt;0.05</b>

Table 2:

<i>Treatment Group</i>	<i>Percent Weight Change (%)</i>	<i>Group Comparison p-value</i>	<i>Fractional Shortening (%)</i>	<b>Group Comparison p-value</b>
<b>Week 6 PI</b>				
A. Catheter/Doxorubicin	-4.13±7.49	A VS.B<0.005 A VS.C=0.01	57.51±7.86	A VS.B=0.05 A VS.C NS
B. Catheter/Saline	14.54±7.17	B VS.D<0.05	71.24±2.51	C VS. D NS
C. Venotomy/Doxorubicin	-18.23±3.22	C VS.D=0.01	67.18±6.57	
D. Venotomy/Saline	6.36±3.33		71.04±2.25	
<b>Kruskal-Wallis p-value</b>	<b>&lt;0.0005</b>		<b>&lt;0.05</b>	
<b>Week 8 PI</b>				
A. Catheter/Doxorubicin	-4.52±4.26	A VS. B NS A VS. C NS	40.91±4.59	A VS. B<0.005 A VS. C<0.05
B. Catheter/Saline	13.30±1.96	B VS. D NS	70.72±2.07	C VS. D<0.01
C. Venotomy/Doxorubicin	-6.28±4.93	C VS. D NS	59.45±10.14	
D. Venotomy/Saline	10.55±2.35		72.54±0.82	
<b>Kruskal-Wallis p-value</b>	<b>0.01</b>		<b>&lt;0.0005</b>	

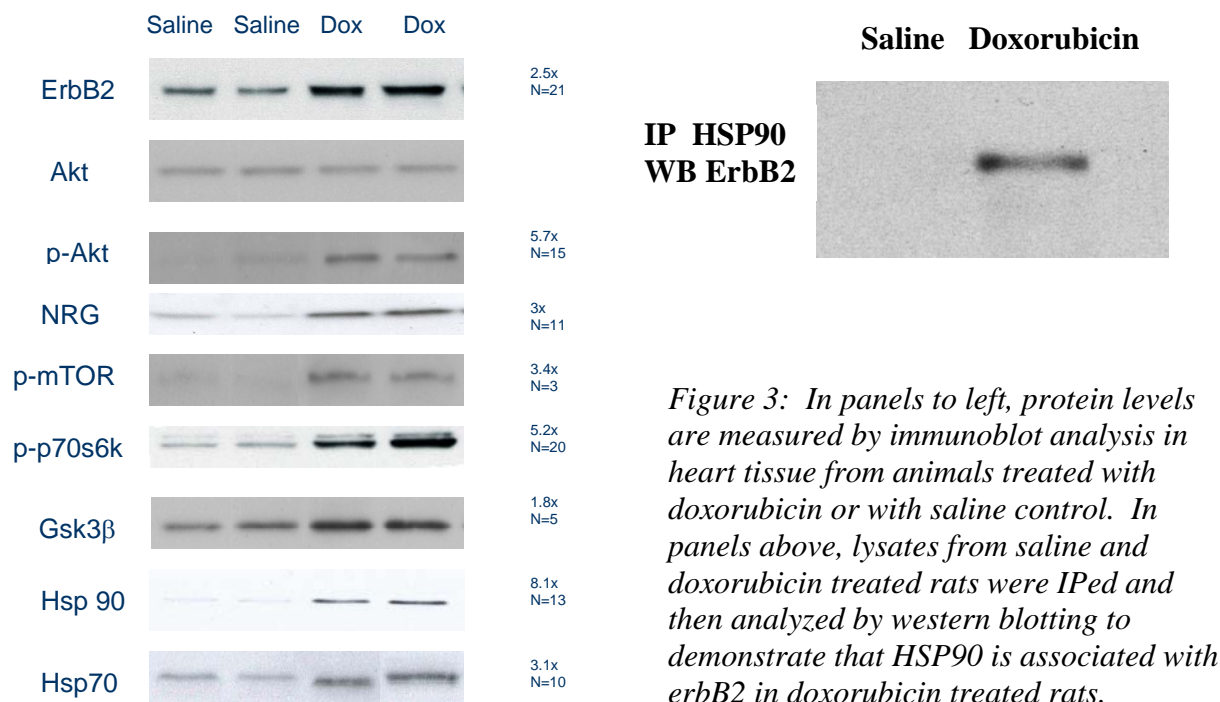
Figure 1:





**Part II: Summary**

Cardiac toxicity is a major drawback to doxorubicin as a cancer-treating drug. Moreover, anti-erbB2 antibody therapy (i.e., trastuzumab) for breast cancer treatment potentiates the effects of doxorubicin on the heart, resulting in even more frequent and severe cardiac toxicity. To understand the mechanism of doxorubicin cardiac toxicity and the synergy of this drug with trastuzumab, we developed an *in vivo* rat model that exhibits progressive loss of cardiac function with doxorubicin treatment. By ten weeks of doxorubicin treatment, rats treated with (2.5 mg/kg dose) doxorubicin (every week for six weeks, frequency) develop a significant reduction in fractional shortening, 39.1% versus 71.1% in saline controls as measured by echocardiography. Immunoblot analysis of cardiac tissue obtained from rats manifesting doxorubicin toxicity showed that this loss of cardiac function is accompanied by a parallel increase in levels of both the erbB2 protein and the major erbB2 ligand, NRG1 $\beta$ . Furthermore, significantly increased levels of the activated (i.e., phosphorylated) forms of AKT, mTOR, p70S6 kinase, and GSK3  $\beta$  are also seen in these doxorubicin-injured hearts, consistent with activation of downstream signaling by the erbB2 receptor during response of the heart to doxorubicin. These elevations of erbB2 protein are not accompanied by elevations in erbB2 mRNA levels, however, suggesting post-transcriptional mechanisms for elevation of erbB2 levels in doxorubicin-injured heart tissue. We therefore considered the potential role of Hsp90, a prototypical cardiac stress response protein, as a stabilizing carrier of erbB2 in the setting of doxorubicin toxicity. Consistent with our expectations, we found significantly increased levels of Hsp90, as well as evidence of Hsp90/ erbB2 binding (by coprecipitation), suggesting that the Hsp90 stress response stabilizes erbB2 and subsequently leads to a cytoprotective response through the AKT signaling pathway. Thus, our results provide a plausible mechanism for the marked susceptibility of the heart to anti-erbB2 therapy in the setting of concomitant doxorubicin therapy.



## **Reportable Outcomes**

### **Manuscripts in press**

**Validation of the Use of Long-Term Indwelling Jugular Catheters in a Rat Model of Cardiac Toxicity**

**Lynn M. Wachtman, Djahida Bedja, Michelle Browning, Scott Pin, Kathy Gabrielson**

### **Manuscripts Submitted**

**erbB2 is increased early in doxorubicin toxicity before cardiac heart function is decreased**

**Kathy Gabrielson, Scott Pin, Djahida Bedja and Nicole Muratore**

## **Presentations**

**AALAS National meeting, St Louis, Missouri, November, 2005 “Overview of Rodent Imaging for phenotyping”**

**ACVP National meeting, “ErbB2 expression in the heart” Boston, Mass. December, 2006**

**Johns Hopkins University ICMIC P50 program December 2005, “Cardiac erbB2- in doxorubicin toxicity -chemotherapy for breast cancer”**

**Cell Stress Society International meeting, March 2006, Concepcion, Chile “Role of Hsp90 in erbB2 expression in heart”**

## **Addendices**

**None to submit**

## Part III summary

### Abstract

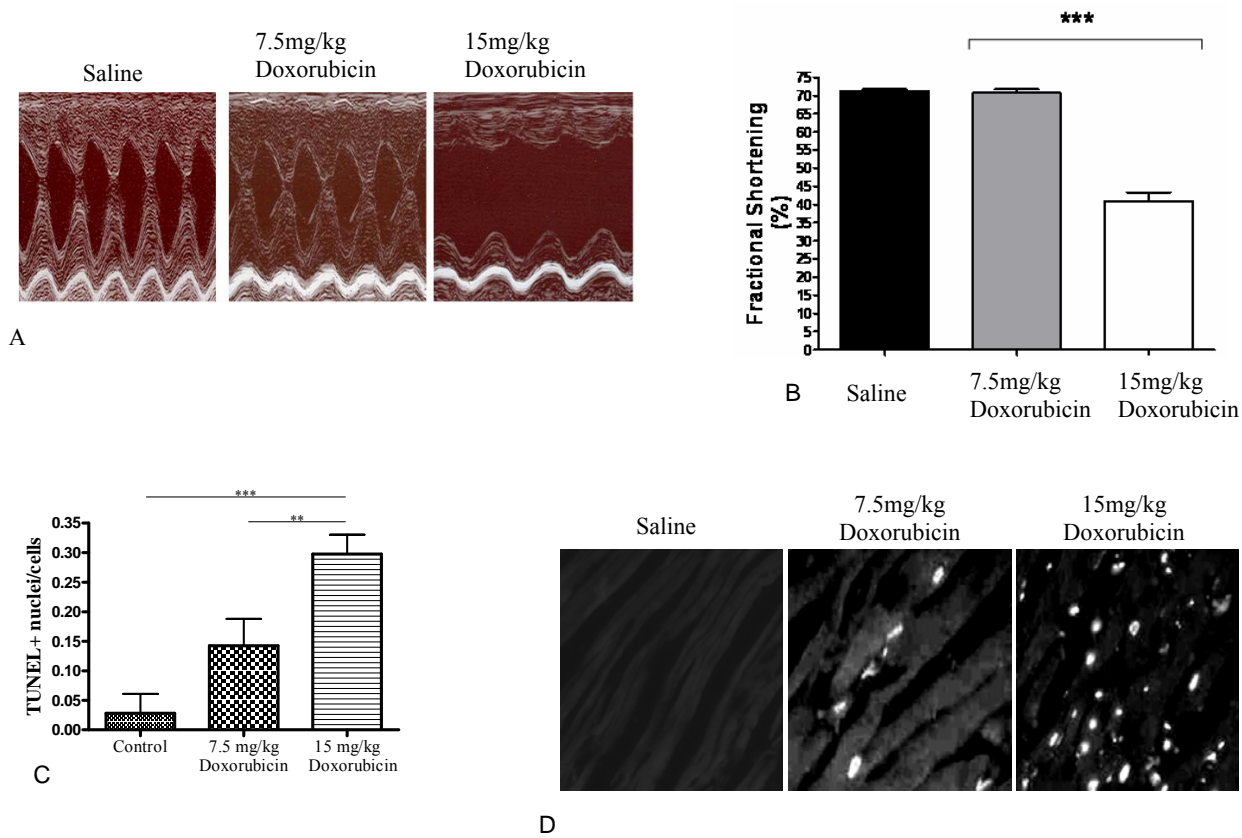
A major drawback to doxorubicin as a cancer-treating drug is cardiac toxicity. To understand the mechanism of doxorubicin cardiac toxicity, and the potent synergic effect seen when doxorubicin is combined with anti-ErbB2 (trastuzumab), we developed an in vivo rat model that exhibits progressive dose-dependent cardiac damage and loss of cardiac function after doxorubicin treatment. The hearts of these animals respond to doxorubicin damage by increasing levels of ErbB2 and the ErbB family ligand, NRG1 $\beta$ , and by activating the downstream Akt signaling pathway. These increases in ErbB2 protein levels are not due to increased ErbB2 mRNA, however, suggesting post-transcriptional mechanisms for regulating this protein in the heart. Accordingly, levels of HSP90, a known ErbB2 protein stabilizer and chaperone, are increased by doxorubicin treatment and co-immunoprecipitation reveals binding of HSP90 to ErbB2. Isolated cardiomyocytes are more susceptible to doxorubicin after treatment with HSP90 inhibitor, 17AAG, suggesting that the HSP90 is protective during doxorubicin treatment. Thus, our results provide one plausible mechanism for the susceptibility of the heart to anti-ErbB2 therapy post-doxorubicin therapy in subclinical and clinical conditions. Additionally, these results suggest that further testing is needed for HSP90 inhibitors under various conditions in the heart.

**Gabrielson, KL**, Bedja, D, Pin S, Gama L, Yuan B, Muratore N, Tsao A., HSP90 and ErbB2 in the Cardiac Response to Doxorubicin Injury, Cancer Research 2007;67(4):1436-41

**Figure 1**

**Effect of doxorubicin on heart function and cardiac cell death.** (A) Representative M-mode echocardiograms of left ventricle from each treatment group. Cumulative dose of 15mg/kg doxorubicin induces contractility deficits at 10 weeks from the initial injection. (B) Fractional shortening % (index of contractility) is summarized in histogram with Mean  $\pm$  SD. Statistical significance ( $p <$ ) is denoted by \*\*\* (0.001) ( $n=10/\text{group}$ ). (C). Histogram of TUNEL positive nuclei with Mean  $\pm$  SD summarized. Rats were euthanized at 10 weeks and hearts were analyzed by TUNEL assay. Apoptotic cells were determined using terminal deoxynucleotidyl transferase (TdT)-mediated dUTP nick end labeling (TUNEL) according to the manufacturer's instructions (*In Situ* Cell Death Detection Kit, Fluorescein, Roche Applied Science) using antigen retrieval methods. TUNEL-positive cells were determined by direct fluorescence of nuclei incorporated dUTP in cardiomyocytes. Total number of cells per 20X field was quantified by H&E staining and TUNEL-positive nuclei were expressed as a percentage of total cells (nuclei) in each heart with 5 fields/heart analyzed. Statistical significance ( $p <$ ) is denoted by \*\* (0.01), \*\*\* (0.001). (D) Representative images of TUNEL fluorescence (converted to black/white) for saline, 7.5 mg/kg and 15 mg/kg Dox treated hearts.

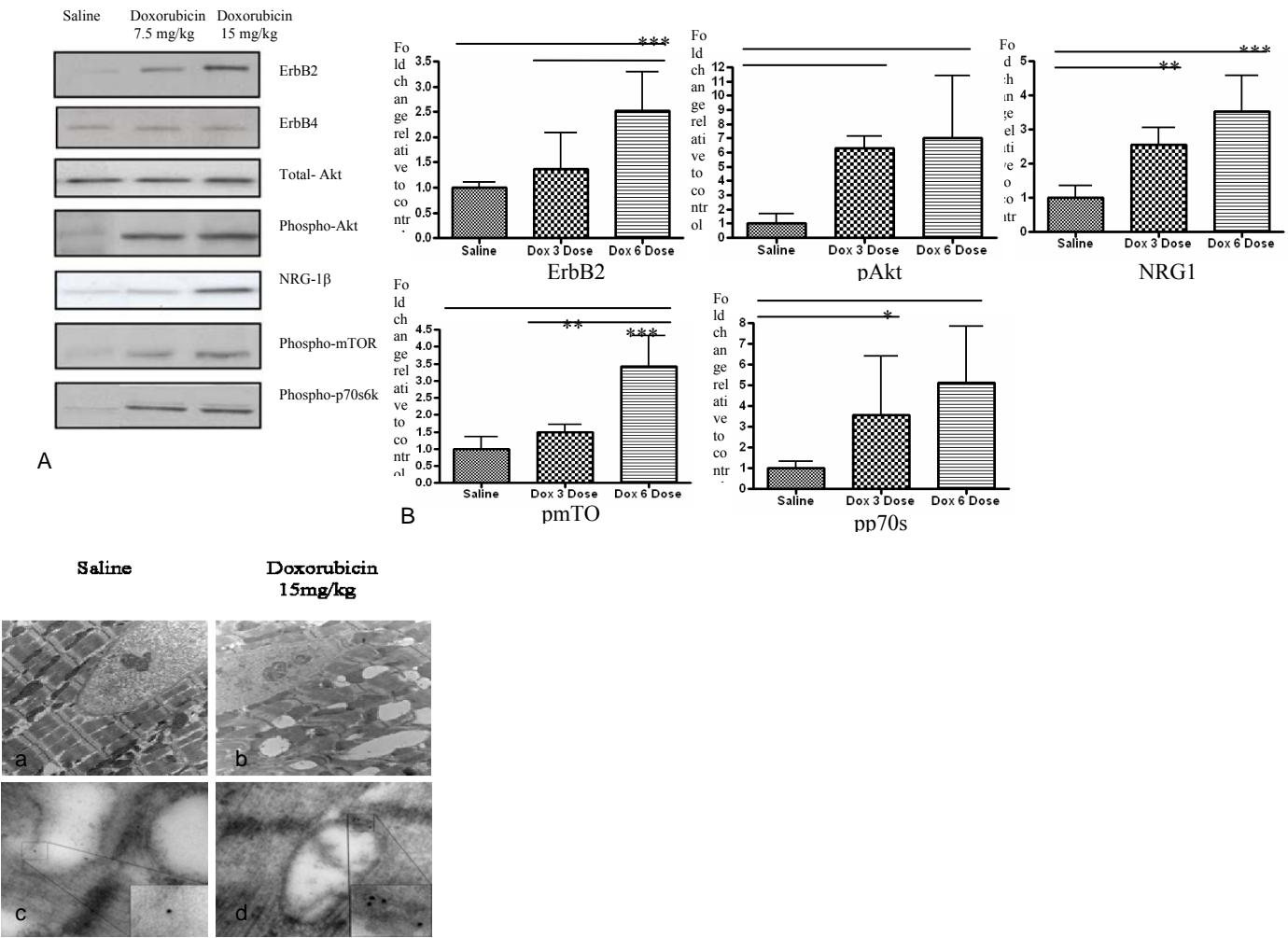
Figure 1





**Figure 2**

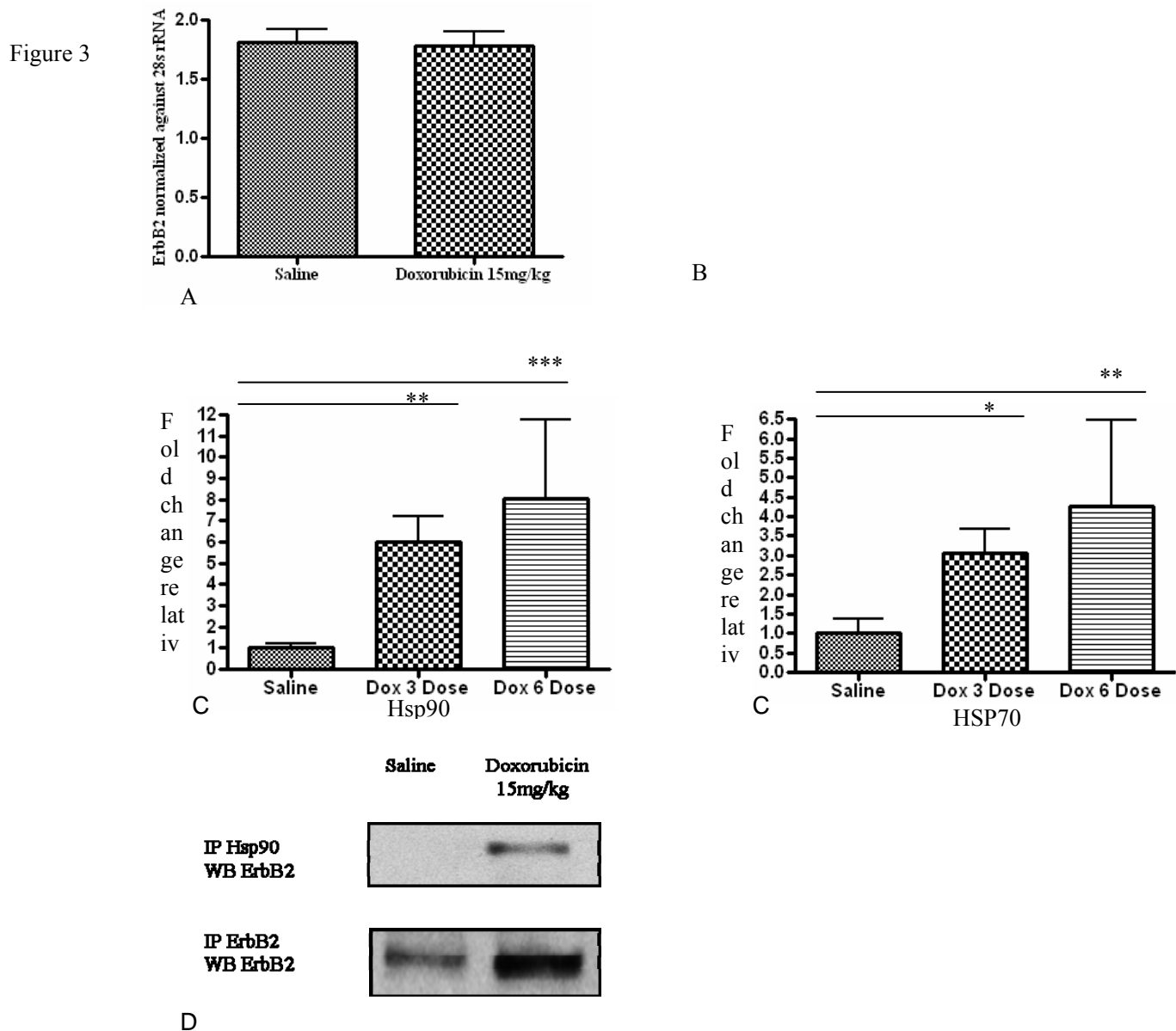
**Effects of doxorubicin on ErbB2 pathway expression in the heart.** Mid-wall left ventricle samples from control or doxorubicin treated rats were collected 10 weeks after the initial injection and proteins isolated for western blotting (n=10/group). (A) Representative western blots indicate increased expression of the ErbB2 and neuregulin1 $\beta$  by doxorubicin treatment. Phosphorylated forms of downstream ErbB2 pathway proteins are increased in both doxorubicin treatment groups compared to saline. (B) Mean  $\pm$  SD summarized in histograms demonstrating relative protein expression (relative to saline) or activation (phosphorylation) normalized to total Akt. Statistical significance (p <) is denoted by \* (0.05), \*\* (0.01), \*\*\* (0.001). (C) Electron micrograph of hearts from a saline (a and c) compared to a 15 mg/kg doxorubicin treated rat (b and d). 10,000 x magnification (a and b) compares a saline treated rat heart (a) to a doxorubicin treated rat with dilated sarcoplasmic reticulum and dilated t-tubules (b) characteristic changes in doxorubicin cardiomyopathy. Bottom two panels (200,000 x magnification) of immunoelectron micrographs of hearts from a saline (c) and doxorubicin (d) treated rat. Increased immunogold labeling of ErbB2 is observed with doxorubicin treatment versus saline with sub-cellular localization of ErbB2 to cardiomyocyte t-tubules adjacent to Z band.



**Figure 3**

**Transcriptional or post-transcriptional mechanisms to increase ErbB2 protein expression in rat heart.** (A) Histogram of ErbB2 RNA expression (Mean  $\pm$  SD) from doxorubicin and saline treated rats demonstrated no significant differences (N=6/group). Total RNA from left ventricle was reverse transcribed using a Superscript II kit (Invitrogen). cDNA was quantitated by Real Time qPCR using ErbB2 probe and primers and conditions described in text. Amplicons were quantitated against standard curves generated by amplification of in vitro transcribed ErbB2 standard RNA. ErbB2 copy number in each sample was normalized by respective ribosomal 28s.

(B) Effect of doxorubicin on heat shock protein expression in the heart. Representative western blotting indicates elevation of the HSP70 and HSP90 normalized to total Akt in doxorubicin treatments. (C) Histogram shows (Mean  $\pm$  SD) relative to saline expression (n=10/group). Statistical significance ( $p <$ ) is denoted by \* (0.05), \*\* (0.01), \*\*\* (0.001). (D) HSP90 and ErbB2 protein:protein association in doxorubicin treatment. Representative western blotting of immunoprecipitation of HSP90 or ErbB2 with ErbB2 western blotting.

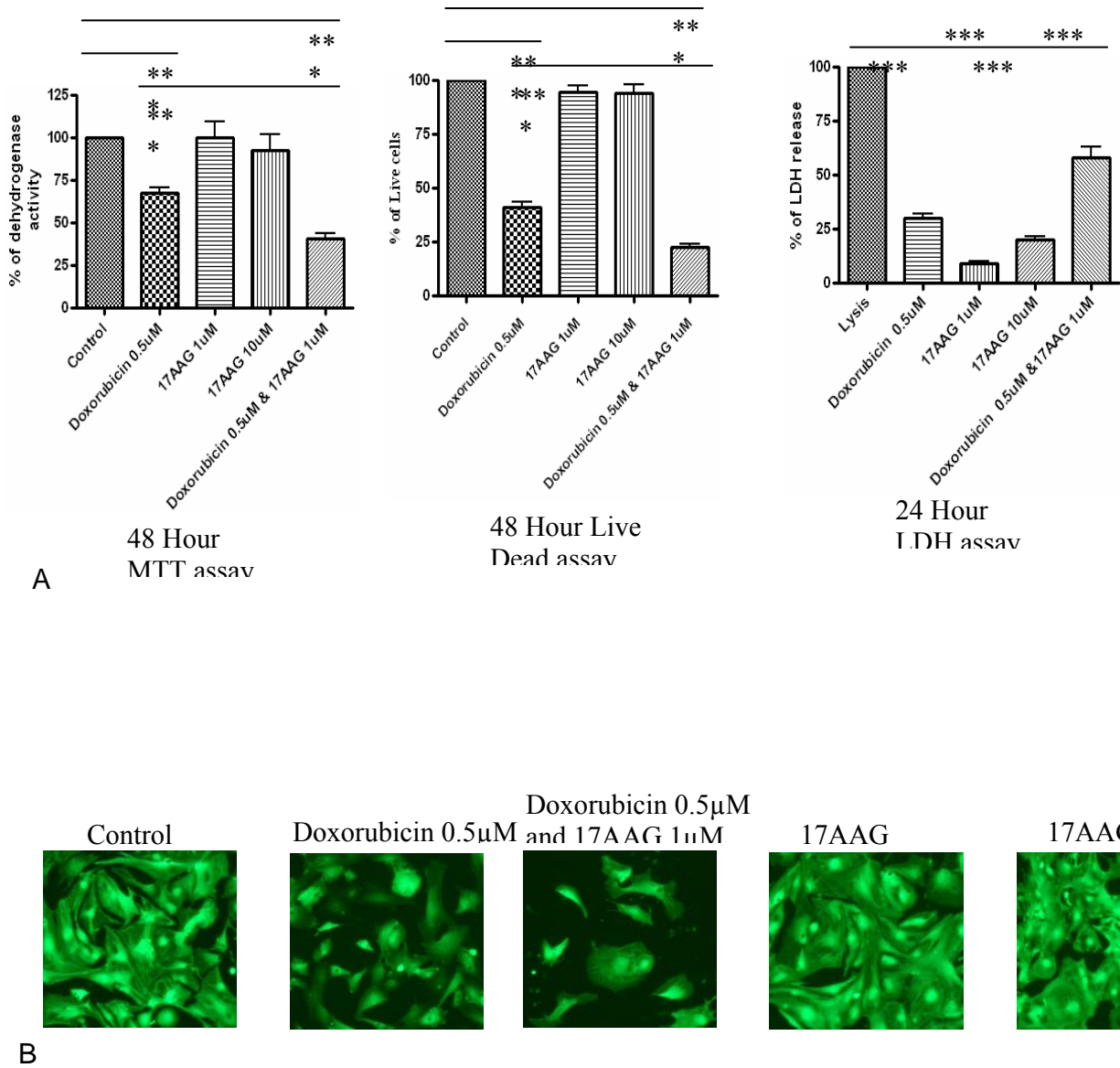


**Figure 4**

**Effect of doxorubicin and HSP90 inhibitor (17AAG) on rat cultured cardiomyocytes.** (A) Cell viability of cardiomyocytes was assessed by LDH, MTT, and Live/dead cell assays on 96 well plates by manufacturer's directions. Histogram shows (Mean  $\pm$  SD) from three separate experiments with 24wells/treatment. 17AAG alone is not toxic at low doses (1  $\mu$ M). However, co-treatment with 17AAG markedly increases the toxicity of doxorubicin. Statistical significance ( $p <$ ) is denoted by \*\*\* (0.001)

(B) Characteristic morphology of cardiomyocytes after 48 hours of treatment before viability assays were performed.

**Figure 4**



## **Reportable Outcomes**

### **Manuscripts in publication or press**

**Gabrielson, KL**, Bedja, D, Pin S, Gama L, Yuan B, Muratore N, Tsao A., HSP90 and ErbB2 in the Cardiac Response to Doxorubicin Injury, Cancer Research 2007;67(4):1436-41

Wachtman, LM., Browning, MD., Bedja, D., Pin, S., **Gabrielson, KL**. Validation of the Use of Long-term Indwelling Jugular Catheters in a Rat Model of Cardiotoxicity. Journal of the American Association for Laboratory Animal Science 2006 September; Volume 45, Number 5: (2006) 55 - 64

Cooper TK, **Gabrielson K.**, Spontaneous Lesions in the Reproductive Tract and Mammary Gland of Female Non-Human Primates, Birth Defects Research Part B: Developmental and Reproductive Toxicology, **in press**

### **National Presentations:**

**ACVP National meeting, “Correlating pathology to in vivo Rodent Imaging” Tuscon, Arizona. December, 2006**

**SOT National meeting, Workshop “In Vivo Imaging for the Toxicologists” Charlotte, NC, march 2007**